SERION ELISA classic

Borrelia burgdorferi IgG/IgM

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SERION ELISA classic Borrelia burgdorferi IgG/IgM

Enzyme Immunoassay for detection of human antibodies (IgG/IgM) For sale in the U.S. for Research Use Only. Not for use in diagnostic procedures.

IgG-Kit (quantitative) order number: ESR121G **IgA-Kit (quantitative**) order number: ESR121M

Tests evaluated: Immunomat TWINsystem / BASEplus, Dade Behring BEP ® III / BEP ® 2000, DSX, manually

1 INTENDED USE

The SERION ELISA *classic* Borrelia burgdorferi IgG and the SERION ELISA *classic* Borrelia burgdorferi IgM are quantitative and qualitative tests for detection of human antibodies in serum, plasma or cerebrospinal fluid directed against Borrelia burgdorferi sensu lato. For sale in the U.S. for Research Use Only. Not for use in diagnostic procedures.

2 BACKGROUND

Borrelia burgdorferi is the infectious agent which causes the disease syndrome known as Lyme-Borreliosis, named after the town Old Lyme in Connecticut (USA). The infectious agent was discovered in 1982 by Willy Burgdorfer and is classified into the taxonomic group of the spirochetes. The bacteria are transmitted to human hosts by infected ticks (Ixodes ricinus, Europe; Ixodes scapularis, USA).

B. burgdorferi sensu stricto, **B.** garinii and **B.** afzelii are the most important human pathogens of the genospecies Borrelia burgdorferi sensu lato. All three are distributed throughout Europe in all temperate climate zones while in the USA human pathogenic strains belong to the species B. burgdorferi sensu stricto. Reservoirs for the bacteria in the wild include a variety of small wild mammals, particularly mice.

The infection rate of adult ticks, nymphs and larvae with Borrelia burgdorferi ranges from 0-50%, dependent upon geographical region and tick populations, while in endemic regions of Germany the infection rate is 10-30 %.

Lyme Disease is a **multisystemic infection** with many possible manifestations. Single or multiple organs may be involved. The time course of Lyme Disease can be divided into three separate stages: **Stage I,** early localized infection, **Stage II,** early disseminated infection, and **Stage III,** late persistent infection.

A European strain of the genospecies Borrelia garinii in addition to the strain PKo has been used as antigen for coating on microtiter wells. The genospecies used expresses the proteins OspC and VlsE and P100. To increase the test's sensitivity, recombinant VlsE is added to the coating antigen. Specificity of the assay is optimized by supplementing the dilution buffer with a treponema phagedenis lysate. The lysate absorbs potentially cross-reactive treponema antibodies.

3 SERION ELISA classic - TEST PRINCIPLE

Microtiter wells are coated with **antigens**. This constitutes the **solid phase**. Sample is added to the wells and any antibodies specific for the antigen present will bind to the solid phase. After removal of unbound material, anti-human **IgG or IgA or IgM conjugated** to an enzyme (**alkaline phosphatase**) is allowed to react with the immune complex. After removal of excess conjugate by washing, an appropriate **substrate** (**paranitrophenylphosphate**) is added, with which the conjugated enzyme reacts producing a **coloured derivative of the substrate**. The colour intensity is proportional to the level of specific antibody bound and can be quantified photometrically.

4 COMPONENTS OF THE KIT

Test components	amount/
	volume
Break apart microtiter test strips each with 8 antigen coated single wells (CV 96) MTP,	12
1 frame	
the coating material is inactivated	
Standard serum (ready-to-use) STD	2 x 2 ml
Human serum in phosphate buffer with protein; negative for anti-HIV-Ab, HBs-Ag ($\underline{\text{H}}$ epatitis $\underline{\text{B}}$ -Virus- $\underline{\text{s}}$ urface antigen) and anti-HCV-Ab; preservative: < 0.1 % sodium azide	
colouring: Amaranth O	
Negative control serum (ready-to-use) NEG	2 ml
Human serum in phosphate buffer with protein; negative for anti-HIV-Ab, HBs-Ag ($\underline{\text{H}}$ epatitis $\underline{\text{B}}$ -Virus- $\underline{\text{s}}$ urface antigen) and anti-HCV-Ab; preservative: < 0.1 % sodium azide	
colouring: Lissamine green V	
Anti-human-IgG-, IgA-, IgM-conjugate (ready-to-use) APC	13 ml
Anti-human-IgG, -IgA, -IgM from goat (polyclonal), conjugated to alkaline phosphatase, stabilized with protein stabilization solution	
preservative: 0.01 % methylisothiazolone, 0.01 % bromnitrodioxane	
Washing solution concentrate (sufficient for 1000 ml) WASH	33.3 ml
Sodium chloride solution with Tween 20, 30 mM Tris	
preservative: < 0.1 % sodium azide	
Dilution buffer S2 (order number: B231-S2) DILBS2	2 x 50 ml
(Special dilution buffer only for SERION ELISA <i>classic</i> Borrelia burgdorferi IgG/IgM. Shake immediately before use!)	
Phosphate buffer with protein and Tween 20; ultrasonicate of Treponema phagedenis	
preservative: < 0.1 % sodium azide	
0.01 g/l Bromphenol blue sodium salt	
Stopping solution STOP	15 ml
1.2 N sodium hydroxide	
Substrate (ready-to-use) pNPP	13 ml
Para-nitrophenylphosphate, solvent free buffer	
preservative: < 0.1 % sodium azide	
(Substrate in unopened bottle may have a slightly yellow color. This does not reduce the quality of the product!)	
Quality control certificate with standard curve and evaluation table INFO	1
(quantification of antibodies in IU/ml or U/ml)	

5 MATERIAL REQUIRED BUT NOT SUPPLIED

- common laboratory equipment
- for the IgM-ELISA: SERION Rf-Absorbent (Order no. Z200/20ml, Z4/4 ml)
- photometer for microtiter plates with filter, wavelength 405 nm, recommended reference wavelength 620 nm 690 nm (e.g. 650 nm)
- incubator 37°C
- moist chamber
- distilled water

6 STORAGE AND STABILITY

Reagent	Storage	Stability	
microtiter strips	unopened	see expiry date on	
(antigen)		microtiter plate	
,	after opening at 2-8°C in closed aluminum bag with	minimum shelf-life	
	desiccant	4 weeks	
	Strips which are not used must be stored in the press-seal bag of aluminum compound foil under dry and airtight conditions!	shelf-life in case of proper use and storage until expiry date	
control sera /	after opening at 2-8°C	until expiry date;	
standard sera		24 months from date of production	
conjugate	ready-to-use solution at 2-8°C	until expiry date	
	Avoid contamination (sterile tips!)	28 months from date of production	
dilution buffer (order number:	(only for SERION ELISA <i>classic</i> Borrelia burgdorferi IgG/IgM)		
B231-S2)	after opening at 2-8°C	24 months	
	Discard cloudy solutions!		
	unopened	until expiry date; 36 months from date of production	
washing solution	concentrate after opening at 2-8°C	until expiry date	
	working dilution at 2-8°C	2 weeks	
	working dilution at room temperature	1 week	
	Bottles used for the working dilution should be cleaned		

	regularly. Discard cloudy solutions.	
substrate	ready-to-use solution at 2-8°C, protected from light!	until expiry date
	Avoid contamination (sterile tips!) Discard when solution turns yellow (extinction against distilled water > 0.25).	24 months from date of production
stopping solution	after opening at room temperature	until expiry date

7 TEST PROCEDURE SERION ELISA classic

7.1 Evidence of deterioration

Only use SERION ELISA *classic* reagents for test procedure, since all reagents are matched. In particular standard and control sera are defined exclusively for the test kit to be used. Do not use them in other lots. Dilution buffer, washing solution and substrate solution can be used for all SERION ELISA *classic* kits irrespective of the lot and the test.

There are three different conjugate concentrations for each immunoglobulin class: LOW, MEDIUM, HIGH

The classic fication is written on each label as follows:

e.g. IgG + lowly concentrated IgG conjugate

IgG ++ medium concentrated IgG conjugate

IgG +++ highly concentrated IgG conjugate

In rare cases the use of special conjugate is necessary to guarantee consistent quality for our products. Special conjugates are produced in a separate lot and do not carry the "+" sign. Therefore, special conjugates are not exchangeable with other conjugates.

Please pay close attention to notifications on labels!

Unopened, all components of the SERION ELISA *classic* kits may be used up to the dates given on the labels, if stored at +2°C to +8°C. Complete stability and storage data are described under "6. Storage and Stability".

Each reagent has been calibrated and optimized for the test. Dilution or alteration of these reagents may result in a loss of sensitivity.

Avoid exposure of reagents to strong light during storage and incubation. Reagents must be tightly closed to avoid evaporation and contamination with microorganisms since incorrect test results could occur due to interference from proteolytic enzymes.

To open the press-seal bag please cut off the top of the marked side, only. Do not use the strips if the aluminum bag is damaged or if the press-seal bag with remaining strips and desiccant was not properly reclosed.

Bring all reagents to room temperature before testing.

<u>Use aseptic techniques</u> for removing aliquots from the reagent tubes to avoid contamination. To avoid false positive results ensure not to contact or sprinkle the topwalls of wells while pipetting conjugate. Take care not to mix the caps of the bottles and/or vials.

Reproducibility is dependent on <u>thorough mixing of the reagents</u>. Shake the flasks containing control sera before use and also all samples after dilution (e.g. by using a vortex mixer).

Be sure to pipette carefully and <u>comply with the given incubation times and temperatures</u>. Significant time differences between pipetting the first and last well of the microtiter plate when filling samples/control sera, conjugate or substrate may result in different "pre incubation" times, which may influence the precision and reproducibility of the results.

Optimum results can only be achieved if SERION ELISA *classic* instructions are followed strictly.

The test is not valid, if the <u>lot-specific validation criteria</u> on the quality control certificate are not fulfilled.

Inadequate washing will affect the test results:

The washing procedure should be carried out carefully. If the washing procedure is carried out automatically follow the instruction manual of the respective washer. Flat bottom wells are used for SERION ELISA *classic*. All wells should be filled with <u>equal volumes</u> of washing buffer. At the end of the procedure ensure that the wells are free of all washing buffer by tapping the inverted microtest plate on a paper towel. <u>Avoid foam!</u> Do not scratch coated wells during washing and aspiration. If using an automated washer, ensure it is operating correctly.

7.2 Sample preparation and storage

Lipaemic, hemolytic or icteric samples should only be tested with reservations although in our testing no negative influence has been found. Obviously contaminated samples (serum or plasma) should not be tested due to the risk of wrong results.

Serum, plasma (EDTA, citrate, heparin) or CSF collected according to standard laboratory methods are suitable samples.

Samples must not be thermally inactivated.

7.2.1 Sample preparation

Before running the test, samples must be diluted in dilution buffer $(V_1 + V_2)$ as follows:

The SERION ELISA *classic* Borrelia burgdorferi IgG/IgM includes a **special dilution buffer Borrelia burgdorferi IgG/IgM** (order number: B231-S2) with lysate of Treponema phagedenis for the absorption of potentially cross-reacting spirochete-antibodies in sera. Shake bottle with dilution buffer Borrelia burgdorferi IgG/IgM immediately before use!

SERION ELISA classic Borrelia burgdorferi IgG

$V_1 + V_2 = 1 + 100$	add	10 μl	sample
	each to	1000 μ1	dilution buffer

After dilution and before pipetting into the microtiter plate the samples must be mixed thoroughly to prepare a homogenous solution.

SERION ELISA classic Borrelia burgdorferi IgM

Rheumatoid factors are **autoantibodies mainly of the IgM-class**, which preferably bind to IgG-immune-complexes. The presence of non-specific IgM-antibodies (rheumatoid factors) can lead to **false-positive** results in the IgM-assay. Furthermore, the possibility exists, that weak-binding pathogen-specific IgM-antibodies are displaced by stronger-binding IgG-antibodies. In this case, IgM-detection can lead to **false-negative** results. Therefore it is necessary to pretreat samples with rheumatoid factor-absorbens prior to IgM detection (SERION Rheumatoid Factor-Absorbent, Order-No. Z4 (4 ml/20 tests), Z200 (20 ml/100 tests)).

Before running the test, rheumatoid factor-absorbent (V_1) must be diluted 1+4 in dilution buffer (V_2) .

$V_1 + V_2 = 1 + 4$	add	200 μ1	Rf-absorbent
	each t	ο 800 μ1	dilution buffer

Samples (V₄) must be diluted in this Rf-dilution buffer (V₃)

$V_4 + V_3 = 1 + 100$	add	10 μl	sample
	each to	1000 μl	Rf-dilution buffer

After dilution and before pipetting into the microtiter plate the samples must be mixed thoroughly to prepare a homogenous solution.

7.2.2 Sample storage

The stoppered samples can be stored in a refrigerator up to 7 days at 2-8°C. Extended storage is possible at \leq -20°C.

Avoid repeated freezing and thawing of samples.

Diluted samples can be stored at 2-8°C for one week.

7.3 Preparation of kit reagents

7.3.1 Microtest strips

Microtest strips in frame are packed with desiccant in an aluminum bag. Take unrequired cavities out of the frame and put them back into the press-seal bag. Close press-seal bag carefully to ensure airtight conditions.

7.3.2 Control sera / standard sera

Control and standard sera are ready-to-use and must not be diluted any further. They can be used directly for the test run.

For each test run and for each test system - independent of the number of microtest strips to be used - control and standard sera must be included. The cut-off-control should be set up in duplicate. With the quantitative tests the standard serum should also be set up in duplicate.

Do not treat control sera with Rf-absorbent.

7.3.3 Anti-human-IgG-, IgM- or IgA-AP-conjugate (ready-to-use)

Conjugates with the same concentration and within the same immunoglobulin class are exchangeable.

Avoid contamination of ready-to-use conjugates (please pour sufficient for test into a secondary container to avoid repeatedly pipetting from the original bottle).

7.3.4 Washing solution

Dilute washing buffer concentrate (V_1) 1:30 with distilled water to a final volume of V_2 .

Example:

buffer concentrate (V ₁)	final volume (V ₂)	
33.3 ml	1000 ml	
1 ml	30 ml	

7.3.5 Dilution buffer for samples (ready-to-use)

Special dilution buffer only for SERION ELISA *classic* Borrelia burgdorferi IgG/IgM (order no. B231-S2).

7.3.6 Substrate (ready-to-use)

For pipetting substrate solution use sterile tips only!

7.3.7 Stopping solution (ready-to-use)

7.4 Overview - test procedure

Borrelia burgdorferi IgG/IgM quantitative

in case of IgM-detection absorption of rheumatoid factor, see No. 7.2.1;

sample dilution¹

1+100

Pipette diluted samples and ready-to-use control sera / standard sera into the microtest wells (100 μ l)

Ú

INCUBATION 60 min./37°C moist chamber

Ų

WASH (4 x 300μ1 DIL WASH)) ²

Û

Pipette conjugate solution APC (100 µl)

Û

INCUBATION 30 min./37°C moist chamber

Û

 $WASH~(4\times300\mu l~\textrm{DIL}~\textrm{WASH}])^{\,2}$

Û

Pipette substrate solution pNPP (100 μl)

Û

INCUBATION 30 min./37°C moist chamber

Û

Pipette stopping solution STOP (100 μl)

Û

READ EXTINCTION AT 405 nm

 1 special dilution buffer for the following test kits: Borrelia IgG & IgM, EBV EA IgG, Parvovirus B19 IgM and Hantavirus Puumala IgG & IgM

²for manual use: at the end of the procedure tap plate on paper towel

7.5 Test procedure

- 1. Place the required number of cavities in the frame and prepare a protocol sheet.
- 2. Add each **100** µl of diluted sample or ready-to-use controls into the appropriate wells of microtest strips. Spare one well for substrate blank, e.g.:

IgG/IgM quantitative		
	<u>, </u>	
well A1	substrate blank	
well B1	negative control	
well C1	standard serum	
well D1	standard serum	
well E1	sample 1	

- 3. Sample incubation for 60 minutes (+/- 5 min) at 37°C (+/- 1°C) in moist chamber
- 4. After incubation **wash** all wells with washing solution (by automated washer or manually):
 - aspirate or shake out the incubation solution
 - fill each well with 300 μl washing solution
 - aspirate or shake out the washing buffer
 - repeat the washing procedure 3 times (altogether 4 times!)
 - dry by tapping the microtest plate on a paper towel
- 5. Addition of conjugate

Add 100 µl of IgG-/IgM-/IgA-conjugate (ready-to-use) to the appropriate well (except substrate blank)

- 6. Conjugate incubation for 30 minutes (+/- 1 min)* at 37°C (+/- 1°C) in moist chamber.
- 7. After incubation **wash** all wells with washing solution (see above)
- 8. Addition of substrate

Add 100 µl substrate solution (ready-to-use) to each well (including well for substrate blank!)

9. Substrate incubation for 30 minutes (+/- 1 min) * at 37°C (+/- 1°C) in moist chamber.

10. Stopping of the reaction

Add 100 µl stopping solution to each well, shake microtest plate gently to mix.

11. Read optical density

Read OD within 60 minutes at 405 nm against substrate blank, reference wave length between 620 nm and 690 nm (e.g. 650 nm).

8 TEST EVALUATION

SERION ELISA classic Borrelia burgdorferi IgG und IgM (quantitative)

8.1 Single-point quantification with the 4PL method

Optimized assignment of extinction signals to quantitative values is guaranteed by using non-linear functions, which adjust a sigmoide curve without any further transformation to OD-values.

Determination of antibody concentrations with the SERION ELISA *classic* is carried out by the **logistic-log-model (4 PL; 4 parameter)** which is ideal for exact curve-fitting. It is based on the formula:

$$OD = A +$$

$$1 + e^{B(C - \text{In conc.})}$$

The parameters A, B, C, and D are representative for the exact shape of the curve:

lower asymptote
 slope of the curve
 turning point
 parameter A
 parameter B
 turning point
 parameter C
 upper asymptote
 parameter D

For each lot the standard curve is evaluated by Institut Virion\Serion GmbH (Würzburg, Germany) in several repeated test runs under optimal conditions. Time consuming and cost intensive construction of the standard curve by the user is not necessary.

For evaluation of antibody concentrations a lot specific standard curve as well as a lot specific evaluation table is included with each test kit. Appropriate evaluation software is available on request.

Please note, that under special working-conditions internal laboratory adaptations of the incubation times could be necessary.

To compensate for normal test variations and also for test run control a standard serum is used in each individual test run. For this control serum a "reference value" with a validity range is determined by the quality control of the producer. Within this range a correct quantification of antibody concentration is ensured. Since the standard serum is not necessarily a positive control, the value of the standard serum may be borderline or negative in some ELISA tests.

8.2 Criteria of validity

- the substrate blank must be OD < 0.25
- the negative control must be negative
- quantitative ELISA: the mean OD-value of the standard serum must be within the validity range, which is given on the lot specific quality control certificate of the kit (after subtraction of the substrate blank!)
- qualitative ELISA: the mean OD-value of the positive control must be within the validity range, which is given on the lot specific quality control certificate of the kit (after subtraction of the substrate blank!)
- the variation of OD-values may not be higher than 20%.

If these criteria are not met, the test is not valid and must be repeated.

8.3 Calculation SERION ELISA *classic* Borrelia burgdorferi IgG/ IgM (quantitativ)

8.3.1 Non-automated evaluation

For the test evaluation a standard curve and an evaluation table are included in the test kit so that the obtained OD-values may be assigned to the corresponding antibody activity. The reference value and the validity range of the standard serum is given on the evaluation table (quality control certificate).

The blank (A1) must be subtracted from <u>all</u> OD-values prior to the evaluation.

Method 1: Qualitative Evaluation

To fix the cut-off ranges please multiply the mean value of the measured standard-OD with the numerical data of the certificate of quality control (see special case formulas), e.g.:

 $OD = 0.502 \times MW (STD)$ with upper cut-off

 $OD = 0.352 \times MW (STD)$ with lower cut-off

If the measured mean absorbance value of the standard serum is 0.64, the range of the cut-off is in between 0.225-0.321.

Method 2:

Continuous determination of antibody activities using the standard curve.

So called *interassay variations* (day to day deviations and laboratory to laboratory deviations) are compensated by multiplication of the current measured value obtained with a sample with the **correction factor F**. This factor is calculated as follows:

OD-reference value (of standard serum)

F = -

OD-current value (of standard serums)

The procedure is necessary to adjust the current level of the test of the user with the lotspecific standard curve.

First, daily deviations have to be corrected by calculating a factor (correction factor F):

- 1. The mean of the two OD-values of the standard serum has to be calculated and checked that it is within the given validity range.
- 2. Calculation of the factor "F": the given reference value is divided by the mean of the extinction of the standard serum:
 - F = reference value extinction standard serum / mean value extinction standard serum.
- 3. All measured values of samples are multiplied by "F".
- 4. Antibody activities in IU/ml or U/ml can be determined from the standard curve with the corrected values.

8.3.2 Automatic test evaluation with SERION *easy base* 4PL-Software/SERION *evaluate-*Software

After input of the 4 parameters and the reference value of the standard serum, antibody activities are calculated online. If the optical density of the standard is out of the valid range, the following message will appear:

SERION easy base 4PL-Software:

"Standards are not in tolerance range" and/or "Distance between standards is greater than 20 %."

SERION *evaluate-***Software**:

"Standard values out of ranges in following groups: Group 1-24. Standard value differ more than 20 % in following groups: Group 1-24."

In these cases the test run is invalid and should be repeated.

Parameters and reference value need to be changed only if there is a change of lot (evaluation table shows parameters and reference values). Correct input of the lot specific data can be checked on the basis of the IU/ml or U/ml assigned to the standard serum. The calculated mean value of the units has to correspond to the unit value indicated on the lot specific certificate. There is an automatic correction of the measured values. In the standard version the printout displays the following:

sample code OD-value IU/ml or U/ml evaluation

9 STATEMENTS OF WARNING

9.1 Statements of warning

The SERION ELISA *classic* is only designed for qualified personnel who are familiar with good laboratory practice.

All kit reagents and human specimens should be handled carefully, using established good laboratory practice.

- This kit contains human blood components. Although all control- and cut-off-sera have been tested and found negative for HBs-Ag-, HCV- and HIV-antibodies, they should be considered potentially infectious.
- Do not pipette by mouth.
- Do not smoke, eat or drink in areas in which specimens or kit reagents are handled.
- Wear disposable gloves, laboratory coat and safety glasses while handling kit reagents or specimens. Wash hands thoroughly afterwards.
- Samples and other potentially infectious material should be decontaminated after the test run.
- Reagents should be stored safely and be unaccessible to unauthorized access e.g. children.
- Stopping solution: corrosive (C); causes acid burn (R34) use safety glasses, gloves and laboratory coat while handling!

9.2 Disposal

Please observe the relevant statutory requirements!

10 BIBLIOGRAPHY

- 1. Aguero-Rosenfeld et al.: Diagnosis of Lyme borreliosis. Clin. Microbiol. Rev. 18 (2005), 484-509
- 2. Goettner G. et al.: Improvement of Lyme borreliosis serodiagnosis by a newly developed recombinant immunoglobulin G (IgG) and IgM line immunoblot assay and addition of VIsE and DbpA homologues; J Clin Microbiol. 2005 Aug; 43 (8): 3602-9
- 3. Liang, F.T. et al.: An Immunodominant Concerved Region within the Variable Domain of VlsE, the Variable Surface Antigen of Borrelia burgdorferi. Journal of Clinical Immunology, 1999, 163: 5566-5573
- 4. Prouqui, P. et al.: Guidelines for the diagnosis of tick-borne bacterial diseases in Europe.clin. Microbiol. Infect.: 10 (2004), 1108-1132
- 5. Stanek, G. et al.: European Union concerted action on risk assessment in Lyme borreliosis: clinical case definitions for Lyme borreliosis. Wien. Klin. Wochenschr.: 108/23 (1996); 741-747
- 6. Tewald, F., Braun, R.
- 7. Durchführung und Interpretation serologischer Tests bei Verdacht auf Borrelieninfektion
- 8. Clin. Lab. 44 (1998); 897-902
- 9. Wilske, B.: Review; Diagnosis of Lyme Borreliosis in Europe. Vector-Borne and Zoonotic Diseases, Vol. 3, No. 4, 2003
- 10. Wilske, B. et al.: MIQ12 Lyme-Borreliose. Qualitätsstandards in der mikrobiologischinfektiologischen Diagnostik. Urban & Fischer Verlag, München Jena 2000, 1-59
- 11. English version: http://nr2-borrelien.lmu.de
- 12. Wilske, B. und Schrifer, M.: Borrelia. Manual of Clinical Microbiology (8th edition). ASM Press, Washington D.C. (2003), 937-954